

WARM STRATIFICATION OF APPLE SEEDS LEADS TO ALTERATIONS IN BIOTIN- CONTAINING PROTEIN LEVELS

M. Tyminski, K. Ciacka, P. Staszek, A. Wal, J. Zak, U. Krasuska

Department of Plant Physiology, Warsaw University of Life Sciences-SGGW,
Nowoursynowska 159, 02-776 Warsaw (Poland)

Abstract

Warm stratification stimulates apple (*Malus domestica* Borkh.) seeds' ageing. In our work, warm stratification was used as a treatment leading to the controlled seeds' deterioration. We examined the levels of biotin-containing proteins (indicators of seeds' viability) and ubiquitin-marked proteins in the embryonic axes of aged seeds. Alterations in the level of biotin-containing proteins depended on the duration of warm stratification. No change in the content of ubiquitinated proteins was observed.

Key words: biotinylated proteins, seeds ageing, ubiquitin-marked proteins, warm stratification

Introduction

Seeds are the main mobile form of Spermatophyte, enabling dispersal in the environment. Seeds' longevity, defined as the total time span during which seeds remain viable, is a significant trait for ecology, agronomy, and economy (Walters et al., 2010, Morscher et al., 2015). Seed quality impacts germination and seedlings' development. Seed viability depends on plant species; e.g. long-term (more than 2,000 years) seed longevity has been described for the date palm (*Phoenix dactylifera* L.) (Sallon et al., 2008). However, prolonged seed storage leads to ageing; even favourable conditions (low humidity and low temperature) do not prevent viability loss (Walters et al., 2010, Morscher et al., 2015). Inappropriate storage conditions accelerate seed deterioration. During seeds' loss of viability, adverse metabolic changes, such as increased membrane permeability and genetic/proteomic damage, occur.

Seeds which tolerate dehydration (orthodox seeds) are more resistant than recalcitrant seeds (sensitive to desiccation) to adverse environmental factors, so they age slowly. Ageing of orthodox seeds can be artificially induced at high temperatures and high relative humidity. Accelerated ageing has been recognized as a good predictor of the loss of seeds' storability (Priestley, 1986). Alterations in the level of specific proteins could be used as markers of seed longevity (Rajjou et al., 2008, Sano et al., 2016). It has been proposed that a group of late embryogenesis abundant (LEA) proteins participate in modulating seed longevity (Sano et al.,

2016). As was demonstrated for pea (*Pisum sativum* L.) seeds, seed-specific biotinylated protein (SBP65, a member of the 3 LEA proteins) is accumulated at the end of the maturation state (Duval et al., 1994a). The decrease of SBP65 was observed during pea seeds' loss of dormancy, pointing to the involvement of these proteins in metabolic processes at early phases of germination (Duval et al., 1994b). Reduction of SBP65 levels was also demonstrated for the embryos of viviparous pea mutant (*vip-1*), which germinate precociously in the pods (Dehaye et al., 1997). It is suggested that this protein is engaged in maintaining seed longevity by scavenging free biotin, which is a cofactor of several carboxylases and decarboxylases (Sano et al., 2016). Nevertheless, there is no information about changes in the levels of biotin-containing proteins in seeds after artificially induced ageing.

In the literature, the mechanism of the degradation of biotin-containing proteins in plant cells is not well described. Proteolysis could be related to ubiquitination – labelling of proteins for degradation in proteasome (Stuttmann et al., 2009). There is no data concerning alteration in ubiquitin-marked protein content in seeds subjected to warm stratification.

Apple (*Malus domestica* Borkh.) seeds belongs to the orthodox type and are characterised by deep dormancy (Lewak, 2011) which can be broken by cold stratification (at 4°C in wetted sand) (Lewak, 2011, Dębska et al., 2013). On the contrary, embryos isolated from seeds subjected to long-term warm stratification (at 25°C in wetted sand) do not germinate well. Moreover, seedlings developed from those embryos have morphological anomalies (Dębska et al., 2013).

The aim of the work was to investigate the levels of biotin-containing proteins and ubiquitin-marked proteins isolated from embryonic axes of warm stratified apple seeds to evaluate loss of vigour during artificially induced ageing. The obtained data provide more insights in the relationship between seed longevity and storage conditions.

Material and methods

Apple (*Malus domestica* Borkh.) seeds were stratified at 32°C for 7, 14, 21 and 40 days. Embryos isolated from stratified seeds were placed on glass Petri dishes containing filter paper wetted with distilled water. Embryos were cultured in a growth chamber at 20°C with 12/12 h (light/dark) photoperiod and light intensity 50 $\mu\text{mol PAR m}^{-2} \text{s}^{-1}$. After 24 h of the culture, embryonic axes were isolated and used for analysis.

Detection of biotin-containing proteins and ubiquitin-marked proteins

Isolation of biotin-containing proteins was done as described by Krasuska et al. (2016). Protein samples (3 μg) were placed on a nitrocellulose membrane. The blocking step was done using 5% (w/v) nonfat dry milk in Tris buffered saline with 0.05% (v/v) Tween-20 (TBST). After incubation the nitrocellulose membrane was washed with TBST and marked with alkaline phosphatase conjugated with streptavidin (S2890 Sigma-Aldrich) solution in 100 mM Tris-HCl pH 9.5 containing

100 mM NaCl and 5 mM $MgCl_2$. Visualization of the spots was performed using nitro blue tetrazolium (NBT) and 5-bromo-4-chloro-3-indolyl phosphate (BCIP).









A similar procedure was done for ubiquitin-marked proteins. After insetting the protein probes on a nitrocellulose membrane and a blocking step, the nitrocellulose was washed with TBST. Then, the nitrocellulose membrane was incubated with a polyclonal anti-ubiquitin antibodies (AS08 307 Agrisera) solution in TBST (1:10,000) for 1 h. After another washing step with TBST, the nitrocellulose membrane was incubated with secondary antibodies (anti-Rabbit-Alkaline Phosphatase conjugated, A3687 Agrisera) 1:30,000. Visualisation of the protein spots was done as described above.

Both detection procedures were done in three independent experiments and the representative results are shown. After a dot blot test, the intensity of the spots was estimated using a densitometry analysis (ImageJ programme).

Results

Densitometry analysis indicated that the content of biotin-containing proteins in axes of apple embryos isolated from seeds stratified for 7 days was 148. A similar level of biotin-containing proteins in homogenates from embryonic axes isolated after 14 days of warm stratification was observed (Fig. 1). Prolonged warm stratification (21 and 40 days) decreased the content of these proteins. Moreover, no significant differences in the intensity of visualised protein spots for these probes were revealed (Fig. 1).

The level of ubiquitin-marked proteins isolated from embryonic axes of seeds stratified for 7, 14, 21 and 40 days was similar (Fig. 1).

Biotin containing proteins					Ubiquitin marked proteins				
Days of warm stratification	7	14	21	40	7	14	21	40	
Dot blot									
Densitometry analysis	148±5	145±12	118±5	120±7	235±10	228±15	240±15	248±13	









Biotin containing proteins					Ubiquitin marked proteins				
Days of warm stratification	7	14	21	40	7	14	21	40	
Dot blot									
Densitometry analysis	148±5	145±12	118±5	120±7	235±10	228±15	240±15	248±13	

Figure 1. The levels of biotin-containing proteins and ubiquitin-marked proteins in axes of apple embryos isolated from seeds stratified at 32°C for 7, 14, 21 or 40 days. Densitometry analysis ± SD.

Discussion

The exposure of seeds to high temperature and high humidity is known as the accelerated ageing procedure and can be performed to indicate seeds' vigor and storability. A rapid deterioration process of seeds points to limited ability to germinate (McDonald, 1999). As was demonstrated for sunflower (*Helianthus annuus* L.) seeds, storage at 45°C and 100% RH reduced germination and led to the growth of atypical seedlings. Finally, such conditions resulted in the seeds' death (Corbineau et al., 1988). In apple seeds, warm stratification, opposite to cold stratification, does not alleviate dormancy (Lewak et al., 2011, Dębska et al., 2013). Furthermore, prolonged warm stratification leads to seeds' ageing and the formation of abnormal seedlings or even their death (Dębska et al., 2013).

Proteins and mRNA are stored in mature seeds as they are sufficient for the first phases of germination (Rajjou et al., 2004). Proteomic analyses done for *Arabidopsis* (*Arabidopsis thaliana* (L.) Heynh.) seeds subjected to controlled deterioration have revealed the linkage between loss in seed vigour and protein alterations in the dry seeds. During germination, the proteome of seeds with low vigor displayed some abnormalities (Rajjou et al., 2008). Rajjou et al. (2008) demonstrated a decreased level of the dehydrin/RAB (Responsive to abscisic acid) group of LEA proteins during seeds' ageing. The authors also proposed that this group of LEA is the most involved in seed longevity; however, they did not exclude other LEA proteins' participation in maintaining seed vigour. Our results indicated that biotin-containing protein levels in apple embryonic axes decrease as the warm stratification period is prolonged. This could point to a link between apple embryo vigour and the amount of such proteins. A lower abundance of biotin-containing proteins was observed during germination of apple embryos (Krasuska et al., 2016) and during pea seeds' loss of dormancy (Duval et al., 1994b), indicating the involvement of these proteins in metabolic pathways during early phases of seed germination. Moreover, the increased level of oxidation of protein homologous to SBP65 isolated from *Arabidopsis* deteriorated seeds has been demonstrated (Rajjou et al., 2008). It has been proposed that the survival of dry, aged *Arabidopsis* seeds can be altered due to the oxidation of biotin-containing proteins (Rajjou et al., 2008). The increase of protein oxidation is commonly linked to stress conditions and elevation of reactive oxygen species (ROS) levels, which are also high in aged tissues (Rajjou et al., 2008, Dębska et al., 2013). During warm stratification of apple seeds, the increase of ROS production and protein carbonylation (type of oxidation) was observed (Dębska et al., 2013). Oxidation of proteins may stimulate their degradation. As we did not observe any changes in ubiquitin-marked proteins' levels, we suspect that the decrease of biotin-containing proteins' levels may be due to their oxidation and proteolysis.

In conclusion, the presented data indicate the relationship between the warm stratification-induced loss of viability of apple seeds and the decrease of seeds' biotin-containing proteins.

Acknowledgments

The work was supported by grant Opus 12 no. 2016/23/B/NZ9/03462 National Science Centre, Poland, given to U.K.

References

1. Corbineau, F., Rudnicki, R.M., Côme, D. (1988) Induction of secondary dormancy in sunflower seeds by high temperature. Possible involvement of ethylene biosynthesis. *Physiologia Plantarum*, 73: 368-373.
2. Dehaye, L., Duval, M., Viguier, D., Yaxley, J., Job, D. (1997) Cloning and expression of the pea gene encoding SBP65, a seed-specific biotinylated protein. *Plant Molecular Biology*, 35: 605-621.
3. Dębska, K., Krasuska, U., Budnicka, K., Bogatek, R., Gniazdowska, A. (2013) Dormancy removal of apple seeds by cold stratification is associated with fluctuation in H_2O_2 , NO production and protein carbonylation level. *Journal of Plant Physiology*, 170: 480-488.
4. Duval, M., DeRose, R.T., Job, C., Faucher, D., Douce, R., Job, D. (1994a) The major biotinyl protein from *Pisum sativum* seeds covalently binds biotin at a novel site. *Plant Molecular Biology*, 26: 265-273.
5. Duval, M., Job, C., Alban, C., Douce, R., Job, D. (1994b) Developmental patterns of free and protein-bound biotin during maturation and germination of seeds of *Pisum sativum*: characterization of a novel seed-specific biotinylated protein. *Biochemical Journal*, 299: 141-150.
6. Krasuska, U., Ciacka, K., Orzechowski, S., Fettke, J., Bogatek, R., Gniazdowska, A. (2016) Modification of the endogenous NO level influences apple embryos dormancy by alterations of nitrated and biotinylated protein patterns. *Planta*, 244: 877-891.
7. Lewak, S. (2011) Metabolic control of embryonic dormancy in apple seed: seven decades of research. *Acta Physiologia Plantarum*, 33: 1-24.
8. McDonald, M.B. (1999) Seed deterioration: physiology, repair and assessment. *Seed Science and Technology*, 27: 177-237.
9. Morscher, F., Kranner, I., Arc, E., Bailly, C., Roach, T. (2015) Glutathione redox state, tocochromanols, fatty acids, antioxidant enzymes and protein carbonylation in sunflower seed embryos associated with after-ripening and ageing. *Annals of Botany*, 116: 669-678.
10. Priestley, D.A. Seed aging: Implications for seed storage and persistence in the soil. Ithaca, New York: Cornell University Press, 1986.
11. Rajjou, L., Gallardo, K., Debeaujon, I., Vandekerckhove, J., Job, C., Job, D. (2004) The effect of alpha-amanitin on the Arabidopsis seed proteome highlights the distinct roles of stored and neosynthesized mRNAs during germination. *Plant Physiology*, 134: 1598-1613.
12. Rajjou, L., Lovigny, Y., Groot, S.P.C., Belghazi, M., Job, C., Job, D. (2008) Proteome-wide characterization of seed aging in Arabidopsis: a comparison between artificial and natural aging protocols. *Plant Physiology*, 148: 620-641.
13. Sallon, S., et al. (2008) Germination, genetics, and growth of an ancient date seed. *Science*, 320: 1464.

14. Sano, N., Rajjou, L., North, H.M., Debeaujon, I., Marion-Poll, A., Seo, M. (2016) Staying alive: molecular aspects of seed longevity. *Plant and Cell Physiology*, 57: 660-674.
15. Stuttmann, J., et al. (2009) COP9 signalosome – and 26S proteasome dependent regulation of SCFTIR1 accumulation in Arabidopsis. *The Journal of Biological Chemistry*, 284: 7920-7930.
16. Walters, C., Ballesteros, D., Vertucci, V.A., (2010) Structural mechanics of seed deterioration: Standing the test of time. *Plant Science*, 179: 565-573.